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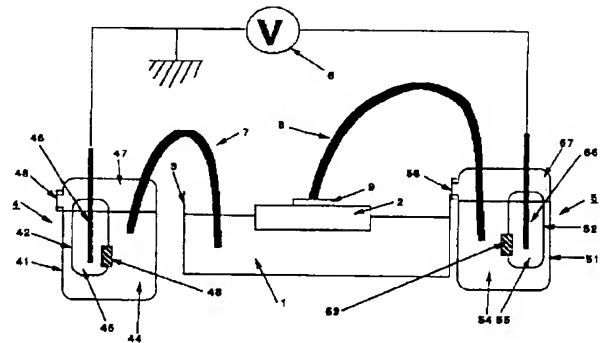
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(54) 【発明の名称】 皮膚表面状態の測定方法及び装置

(57) 【要約】

【課題】 皮膚の表面状態を測定する方法と装置を提供することにあり、特に、皮膚表面への刺激がもたらすヒトの感覚を測定して数値化すること、さらに、感覚がヒトにとって精神的にマイナスである場合に、それを防御する薬剤の効果を測定するための方法とそれらに用いる装置を提供すること。

【解決手段】 容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定する。皮膚表面に刺激を与えてまたは培養液に刺激物質を添加して、前記電位差を測定し、皮膚の刺激度を測定する。皮膚表面に刺激を与えてまたは培養液に刺激物質を添加して、さらに皮膚切片に薬剤を塗布して前記電位差を測定し、薬剤による皮膚刺激の抑制度を測定する。皮膚表面に刺激を与えてまたは培養液に刺激物質を添加して、さらに培養液に薬剤を添加して前記電位差を測定し、薬剤による皮膚刺激の抑制度を測定する。



## 【特許請求の範囲】

【請求項 1】 容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定することを特徴とする皮膚表面状態の測定方法。

【請求項 2】 前記培養液が、生理食塩水又はアミノ酸類を含む培養液である請求項 1 記載の皮膚表面状態の測定方法。

【請求項 3】 容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、皮膚表面に刺激を与えて電位差を測定し、その値と、刺激を与えない場合の電位差との差から、皮膚の刺激度を測定する方法。

【請求項 4】 前記刺激が、刺激物質によることを特徴とする請求項 3 記載の皮膚の刺激度を測定する方法。

【請求項 5】 前記刺激が、物理的刺激である請求項 3 記載の皮膚の刺激度を測定する方法。

【請求項 6】 容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、培養液に刺激物質を添加して電位差を測定し、その値と、刺激物質を添加しない場合の電位差との差から、皮膚の刺激度を測定する方法。

【請求項 7】 前記刺激物質が、皮膚に対するかゆみ発生物質である請求項 4 または 6 記載の皮膚の刺激度を測定する方法。

【請求項 8】 前記培養液が、生理食塩水又はアミノ酸類を含む培養液である請求項 3 乃至 7 のいずれか一項に記載の皮膚の刺激度を測定する方法。

【請求項 9】 容器内に収容した培養液の液面に表面を上にして刺激を与えた皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、皮膚表面に刺激を与える前あるいは後に、その皮膚表面に薬剤を塗布して電位差を測定し、その値と、薬剤を塗布しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定する方法。

【請求項 10】 容器内に収容した培養液の液面に表面を上にして刺激を与えた皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、皮膚表面に刺激を与える前あるいは後に、培養液に薬剤を添加して電位差を測定し、その値と、薬剤を添加しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定する方法。

【請求項 11】 前記刺激が、刺激物質によることを特徴とする請求項 9 または 10 記載の皮膚刺激の抑制度を測定する方法。

【請求項 12】 前記刺激が、物理的刺激である請求項 9 または 10 記載の皮膚刺激の抑制度を測定する方法。

【請求項 13】 容器内に収容した刺激物質を添加した

培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、培養液に刺激物質を添加する前あるいは後に、その皮膚表面に薬剤を塗布して電位差を測定し、その値と、薬剤を塗布しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定する方法。

【請求項 14】 容器内に収容した刺激物質を添加した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、培養液に刺激物質を添加する前あるいは後に、培養液に薬剤を添加して電位差を測定し、その値と、刺激物質を添加しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定する方法。

【請求項 15】 前記刺激物質が、皮膚に対するかゆみ発生物質である請求項 11 または 13 または 14 記載の皮膚刺激の抑制度を測定する方法。

【請求項 16】 前記培養液が、生理食塩水又はアミノ酸類を含む培養液である請求項 9 乃至 15 のいずれか一項に記載の皮膚刺激の抑制度を測定する方法。

【請求項 17】 培養液と、その液面に表面を上にして浮かべた皮膚切片を収容した容器と、前記培養液に連絡される標準電極部と、前記皮膚切片の皮膚表面に連絡される測定電極部と、前記標準電極部と前記測定電極部との電位差を検出する電位差検出部とを備えたことを特徴とする皮膚表面状態測定装置。

【請求項 18】 前記標準電極部が、標準電極が浸漬された内部液を収容した内チューブと、この内チューブの中に収納し二重チューブを形成する、内部液を収容した外チューブとを備え、前記内チューブ壁には液絡が被着されてなることを特徴とする標準電極部である請求項 17 記載の皮膚表面状態測定装置。

【請求項 19】 前記培養液と標準電極部との連絡が、前記培養液と、前記外チューブ内の内部液との間に結ばれた塩橋によることを特徴とする請求項 18 記載の皮膚表面状態測定装置。

【請求項 20】 前記測定電極部が、測定電極が浸漬された内部液を収容した内チューブと、この内チューブの中に収納し二重チューブを形成する、内部液を収容した外チューブとを備え、前記内チューブ壁には液絡が被着されてなることを特徴とする測定電極部である請求項 17 乃至 19 のいずれか一項に記載の皮膚表面状態測定装置。

【請求項 21】 前記皮膚切片の皮膚表面と測定電極部との連絡が、前記皮膚切片の皮膚表面と、前記外チューブ内の内部液との間に結ばれた塩橋によることを特徴とする請求項 20 記載の皮膚表面状態測定装置。

【請求項 22】 前記培養液が、生理食塩水又はアミノ酸類を含む培養液である請求項 17 乃至 21 のいずれか一項に記載の皮膚表面状態測定装置。

【発明の詳細な説明】

## 【0001】

【発明の属する技術分野】本発明は、皮膚表面の状態を測定する方法及びそれらを測定する装置に関する。さらに詳しくは、培養液の液面に表面を上にして皮膚切片を浮かべた皮膚モデルを作成し、その皮膚表面と培養液との電位差から皮膚表面の状態を測定する方法、また、皮膚表面あるいは培養液に刺激要因を与えて変化する電位差から刺激度を測定する方法、さらにこれらの刺激皮膚に薬剤を加えた時の電位差の変化から皮膚刺激の抑制度を測定する方法、並びにこれらの方法を実施するに適した装置に関する。

## 【0002】

【従来の技術】ヒトの皮膚表面は自然界からさまざまな刺激を受けている。刺激を受けた場合、皮膚に症状として現れると同時にかゆみ、いたみ、ひりつき等のようなヒトの感覚で表現されるものが生じる場合が多く、また、症状が現れず感覚のみのものも多くある。これらの刺激には快適なものもあるが、不快なものも多く、例えばかゆみ等の刺激は、ヒトにとって不快なものである。これらの刺激はヒトの感覚の範疇であり、他人には分からない厄介なものでもある。

【0003】しかしながら、従来皮膚の表面状態を測定する方法がなかったため感覚の度合をヒトの訴えに頼らざるを得なかった。したがって、そのような刺激を客観性のある数値等に置き換えられれば主観的な感覚の共有化ができ、さらに上記数値等を使えば不快な刺激の抑制薬剤の開発速度が急転することが期待される。

## 【0004】

【発明が解決しようとする課題】本発明は上記事情に鑑みてなされたもので、その目的は、皮膚の表面状態を測定する方法と装置を提供することにある。特に、皮膚表面への刺激がもたらすヒトの感覚を測定して数値化すること、さらに、感覚がヒトにとって精神的にマイナスである場合に、それを防御する薬剤の効果を測定するための方法とそれらに用いる装置を提供することにある。

## 【0005】

【課題を解決するための手段】本発明者らは、皮膚表面の電位に注目することにより、上記課題を解決しようと試みた。そして、培養液の液面に表面を上にして皮膚切片を浮かべた簡便な皮膚モデルを作成し、皮膚表面と培養液との電位差を簡便に測定できる方法を見出しこれを活用することにより上記課題を解決した。

【0006】すなわち、本発明による皮膚表面状態測定方法は、容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定することを特徴とする。

【0007】また、容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定し、その値と、刺激を

与えない場合の電位差との差から、皮膚の刺激度を測定することを特徴とする。

【0008】本発明の皮膚刺激度の測定において、前記刺激は、刺激物質または物理的刺激によることができる。

【0009】また、容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、培養液に刺激物質を添加して電位差を測定し、その値と、刺激物質を添加しない場合の電位差との差から、皮膚の刺激度を測定することを特徴とする。

【0010】これら、前記皮膚の刺激度の測定に当たって用いられる刺激物質で最も典型的なものはかゆみ発生物質である。

【0011】また、容器内に収容した培養液の液面に表面を上にして刺激を与えた皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、皮膚表面に刺激を与える前あるいは後に、その皮膚表面に薬剤を塗布して電位差を測定し、その値と、薬剤を塗布しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定することを特徴とする。

【0012】さらに、容器内に収容した培養液の液面に表面を上にして刺激を与えた皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、皮膚表面に刺激を与える前あるいは後に、培養液に薬剤を添加して電位差を測定し、その値と、薬剤を添加しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定することを特徴とする。

【0013】本発明の皮膚刺激抑制度の測定において、前記刺激は、刺激物質または物理的刺激によることができる。

【0014】また、容器内に収容した刺激物質を添加した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、培養液に刺激物質を添加する前あるいは後に、その皮膚表面に薬剤を塗布して電位差を測定し、その値と、薬剤を塗布しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定することを特徴とする。

【0015】さらに、容器内に収容した刺激物質を添加した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差を測定するに当たって、培養液に刺激物質を添加する前あるいは後に、培養液に薬剤を添加して電位差を測定し、その値と、刺激物質を添加しない場合の電位差との差から、薬剤による皮膚刺激の抑制度を測定することを特徴とする。

【0016】前記皮膚刺激の抑制度の測定に当たって用いられる刺激物質で最も典型的なものはかゆみ発生物質である。

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【0017】このような皮膚表面状態測定方法を実施するための装置は、培養液と、その液面に表面を上にして浮かべた皮膚切片を収容した容器と、前記培養液に連絡される標準電極部と、前記皮膚切の皮膚表面に連絡される測定電極部と、前記標準電極部と前記測定電極部との電位差を検出する電位差検出部とを備えたことを特徴とする。

【0018】本発明の皮膚表面状態測定装置に用いられる好ましい標準電極部は、標準電極が浸漬された内部液を収容した内チューブと、この内チューブを中に収納し二重チューブを形成する、内部液を収容した外チューブとを備え、前記内チューブ壁には液絡が被着されてなることを特徴とする標準電極部である。

【0019】この場合の前記培養液と標準電極部との連絡は、前記培養液と、前記外チューブ内の内部液との間に結ばれた塩橋による。

【0020】また、本発明の皮膚表面状態測定装置に用いられる前記測定電極部としては、測定電極が浸漬された内部液を収容した内チューブと、この内チューブを中に収納し二重チューブを形成する、内部液を収容した外チューブとを備え、前記内チューブ壁には液絡が被着されてなることを特徴とする測定電極部を用いることができる。

【0021】この場合の前記皮膚切片の皮膚表面と測定電極部との連絡は、前記皮膚切片の皮膚表面と、前記外チューブ内の内部液との間に結ばれた塩橋による。

【0022】なお、前記各方法及び装置で用いる培養液は、生理食塩水又はアミノ酸類を含む培養液であることが好ましい。

【0023】前記のような本発明による皮膚表面状態測定方法と装置によれば、皮膚切片の皮膚表面と皮膚切片を浮かべた培養液との電位差を好適な装置を用いて測定することにより、皮膚表面の状態を簡便に測定することができる。また、皮膚表面あるいは培養液に刺激要因を与えて前記電位差の変化を測定することにより、ヒトを直接使用することなく、しかもヒトの主観を越えて、刺激を客観的に表すことができる。さらに、薬剤添加による前記電位差の変化から皮膚刺激に対して有効な薬剤の探索に活用でき、新薬剤探索に対して大幅な効果を図ることができる、非常に有用である。

【0024】なお、本発明において、前記皮膚表面状態とは、皮膚表面で起こるさまざまな状態変化をも含めた概念であり、例えば刺激、刺激の抑制等による状態変化及びその度合をも含めたものである。

【0025】

【発明の実施の形態】以下、本発明の方法、本発明の装置の順に説明する。

【0026】本発明の第一の方法は、容器内に収容した培養液の液面に表面を上にして皮膚切片を浮かべ、この皮膚切片の皮膚表面と前記培養液との電位差 (mV) を

測定する。これにより、皮膚の表面状態が測定される。

【0027】本発明の第二の方法は、第一方法の皮膚表面状態の測定に当たって、さらに皮膚切片と培養液からなる皮膚モデルに刺激を与えて前記電位差を測定する。この電位差は、刺激を加えることによって変化する。この変化は表皮の中にイオンの流れが発生したことを意味する。刺激によってイオンの流れの方向が異なる。未処理の皮膚の値に比べて変化があれば、表皮にイオンの動きを発生させる刺激が与えられたと解釈する。その変化値から皮膚の刺激度が測定される。刺激を与えるに当たっては、その場所は皮膚表面でも培養液でもいづれでもよい。皮膚表面の場合は、刺激の与え方として、刺激物質の皮膚への塗布、物理的な刺激等がとられる。また、培養液の場合は刺激物質の添加の方法がとられる。これらの刺激物質としては、神経ペプチド、ヒスタミン、界面活性剤、有機酸、カプサイシン等が挙げられる。これらの中、神経ペプチドとしては、サブスタンス P 等がある。本発明においては、刺激物質がかゆみ発生物質であることが特に有用である。また、前記物理的刺激的例としては、セロテープ (登録商標) によるバリアー破壊等による例等がある。

【0028】本発明の第三の方法は、第二の方法で刺激を与えた皮膚の薬剤添加による抑制度を測定する方法である。本発明においては、前記皮膚モデル系に刺激を与えた後、薬剤を添加して前記電位差を測定し、添加によって変化する電位差の値から薬剤の皮膚刺激の抑制度を測定するものである。

【0029】薬剤の添加方法によって本方法はいろいろな方法をとることができる。すなわち、

(1) ; 皮膚表面に刺激を与えた皮膚切片を表面を上にして培養液に浮かべ、皮膚刺激を与える前あるいは後に皮膚表面に薬剤を塗布して電位差を測定し、薬剤塗布によって変化する電位差の値から薬剤の皮膚刺激の抑制度を測定する。(2) ; (1) の皮膚表面に薬剤を塗布する代わりに培養液に薬剤を添加して電位差を測定し、培養液への薬剤添加によって変化する電位差の値から薬剤の皮膚刺激の抑制度を測定する。(3) ; 刺激物質を添加した培養液に皮膚切片を表面を上にして浮かべ、刺激物質を添加する前あるいは後に皮膚表面に薬剤を塗布して電位差を測定し、薬剤塗布によって変化する電位差の値から薬剤の皮膚刺激の抑制度を測定する。(4) ;

(3) の皮膚表面に薬剤を塗布する代わりに培養液に薬剤を添加して電位差を測定し、培養液への薬剤添加によって変化する電位差の値から薬剤の皮膚刺激の抑制度を測定する。

【0030】また、前記薬剤の塗布は、刺激を与える前でも後でも構わない。薬剤の塗布が刺激を与えた後であれば、刺激を緩和する効果として測定でき、また刺激を与える前であれば、刺激を未然に防止する効果として測定できる。

【0031】第一乃至第三の測定方法において用いられる培養液は、生理食塩水、アミノ酸類などを含む水溶液等が用いられる。また、測定に用いる皮膚切片は、培養皮膚、成形手術等の際得られるヒト皮膚、ヘアレスマウス等の動物由来の皮膚等を用いることができる。

【0032】ヒトの皮膚が刺激を受けた場合、皮膚に症状として現れると同時にかゆみ、いたみ、ひりつき等のようなヒトの感覚で表現されるものが生じる場合が多く、また、症状が現れず感覚のみのものも多くある。本発明の前記第一乃至第三の方法によれば、このような主観的な感覚が客観性のある数値として表すことができるので、例えばヒトを使うことなく刺激を緩和あるいは抑制する薬剤の探索に活用することができ、新薬剤の開発に大いに貢献することができる。

【0033】次に、本発明の方法に好ましく用いられる装置について、図面を参照しながら説明する。

【0034】図1は、本発明における皮膚表面状態測定装置の例を示すものである。図1に示す通り、皮膚表面状態測定装置は、培養液1と、その液面に表面を上にして浮かべた皮膚切片2を収容した容器3と、前記培養液1に連絡される標準電極部4と、前記皮膚切片2の皮膚表面に連絡される測定電極部5と、前記標準電極部4と前記測定電極部5との電位差を検出する電位差検出部6とを有する。

【0035】標準電極部4は、プラスチック製の二重の外チューブ41及び内チューブ42を有し、内チューブ42のチューブ壁には溶液は通過させないが電子もしくはイオンは通過させる機能を持った多孔質セラミックからなる液絡43を有している。この二重の外チューブ41及び内チューブ42の内部には、所定濃度のKC1溶液からなる内部液44、45がそれぞれ満たされており、内チューブ42の内部液45に銀-塩化銀からなる標準電極46が浸漬されている。この標準電極46は、二重の外チューブ41及び内チューブ42の上端を閉じるキャップを兼ねた電極ホルダ47に取り付けられ、そこから導体を介して吊り下げられた状態で内チューブ42内の内部液45に浸漬されている。外チューブ41の上端寄りの周面には、その外チューブ41内に内部液44を補充する液補充口48が設けられている。

【0036】なお、図1の標準電極部4において、二重の外チューブ41及び内チューブ42はいずれもガラス製を使用することもできる。また、液絡43はゼラチン、寒天等で擬晶構造をもたせた溶液など公知のものや多孔質のバイコールガラス等塩橋として用いられるものも含めて使用することができる。また、前記標準電極46はカロメルからなる電極を用いても構わない。また、内部液45にはKC1溶液以外に、塩化ナトリウム、塩化リチウム等の塩化物の溶液が使用できる。外チューブ41内の内部液44は、前記内チューブ42内の内部液45に用いられる塩化物以外に生理食塩水でもよい。

【0037】測定電極部5は、前記標準電極部4と同様の構造を有している。すなわち、プラスチック製の二重の外チューブ51及び内チューブ52を有し、内チューブ52のチューブ壁には多孔質セラミックからなる液絡53を有している。この二重の外チューブ51及び内チューブ52の内部には、所定濃度のKC1溶液からなる内部液54、55がそれぞれ満たされており、内チューブ52の内部液55に銀-塩化銀からなる測定電極56が浸漬されている。この測定電極56は、二重の外チューブ51及び内チューブ52の上端を閉じるキャップを兼ねた電極ホルダ57に取り付けられ、そこから導体を介して吊り下げられた状態で内チューブ52内の内部液55に浸漬されている。外チューブ51の上端寄りの周面には、その外チューブ51内に内部液54を補充する液補充口58が設けられている。

【0038】なお、図1の測定電極部5においても、前記標準電極部4と同様の実施態様をとることができる。すなわち、二重の外チューブ51及び内チューブ52はいずれもガラス製を使用することもできる。また、液絡53は多孔質のバイコールガラスを用いることもできる。また、前記測定電極56はカロメルからなる電極を用いても構わない。また、内部液55にはKC1溶液以外に、塩化ナトリウム、塩化リチウム等の塩化物の溶液が使用できる。外チューブ51内の内部液54は、前記内部液55に用いられる塩化物以外に生理食塩水でもよい。

【0039】さらに、図1における標準電極部4は、その外チューブ41内の内部液44に浸漬された柔軟性のあるプラスチックチューブを生理食塩水を含むアガロースで満たした塩橋7を介してDMEM (Dulbecco's modified Eagle's medium) からなる培養液1と連絡される。このDMEMはアミノ酸類を含んだ公知の培養液であり、市販品として入手できる。この他、生理食塩水を含んだもの等公知のものを用いることもできる。また、塩橋7は公知のものでよく、例えばシリコン等のゴムチューブ等も使用できる。

【0040】また、測定電極部5は、その外チューブ51内の内部液54に浸漬された柔軟性のあるプラスチックチューブを生理食塩水を含むアガロースで満たした塩橋8を介して皮膚切片2の皮膚表面と表面上に置かれたろ紙9を通じて連絡されている。このろ紙9は塩橋8の皮膚切片2の皮膚表面への接触を均一にするためである。なお、塩橋8は公知のものでよく、例えばシリコン等のゴムチューブ等も使用できる。

【0041】前記標準電極部4の標準電極46と測定電極部5の測定電極56は、電位差検出部6に接続され、その間の電位差が測定される。

【0042】なお、皮膚表面状態測定装置における標準電極部として、図1の標準電極部4の外チューブ41のチューブ壁に液絡を設けたものを用いることができる。

外チューブには、図 1 と同様内部液を補充する液補充口を設けてもよい。

【0043】この標準電極部は、図 1 における皮膚表面状態測定装置の標準電極部 4 と同様の機能をもって、図 1 の標準電極部 4 に代えて使用できる。ただし、培養液との連絡は、この標準電極部が培養液に浸漬され、外チューブ壁に設けた液路を介して行われる。なお、この標準電極部における実施態様は図 1 の場合と同様である。

【0044】さらに、皮膚表面状態測定装置における標準電極部として、図 1 の標準電極部 4 の内チューブ 4 2 の部分のみ、すなわち、内チューブ 4 2、液路 4 3、内部液 4 5、標準電極 4 6 のみからなっているものを用いることができる。

【0045】この標準電極部も、図 1 における皮膚表面状態測定装置の標準電極部 4 と同様の機能をもって、図 1 の標準電極部 4 に代えて使用できる。ただし、培養液との連絡は、この標準電極部が培養液に浸漬され、チューブ壁に設けた液路を介して行われる。なお、この標準電極部における実施態様も図 1 の場合と同様である。

【0046】

【実施例】次に、本発明のより具体的な実施例について説明する。

【0047】（実施例 1）図 1 に示した皮膚表面状態測定装置を使用し、培養液に対する皮膚表面の電位差（mV）を測定した。測定に用いた皮膚サンプルは、ヘアレスマウスの皮膚切片を用い、（1）皮膚表面に何も処置を施さないもの、（2）刺激物質であるサブスタンス P（SP）を培養液に添加したもの、（3）SP とともに、刺激の防御物質として知られている SP アンタゴニスト（[D-Arg<sup>1</sup>, D-Trp<sup>7,9</sup>, Leu<sup>11</sup>]-Substance P）を培養液に添加したもの、の三種類を使用した。結果を図 2 に示す。

【0048】図 2 において、横軸は SP を添加してからの経過時間を示す。また、SP アンタゴニストを添加した場合は、それを加えた時点をゼロとなるようにされる。縦軸は電位差を示す。測定値は 5 回測定した平均で表し、そのばらつきを同時に示してある。

【0049】図 2 に示されたように、皮膚表面に何も処置を施さないもの（コントロール）に対して、SP を培養液に添加すると、電位差の急激な変化が見られる。添加後 1 時間をピークとし電位差の大きな変化の状態が続く、そして 2 時間後にコントロールに近い値にもどっている。一方、SP を添加する前に SP アンタゴニストを培養液に添加しておく、と、電位差の急激で大きな変化が見られることなく、添加後 2 時間まではほぼコントロールと同じ値を示していることが分かる。このことから、SP とともに SP アンタゴニストを添加した場合、SP アンタゴニストが SP による刺激を抑制し、電位差変化を抑制することが分かる。

【0050】（実施例 2）図 1 に示した皮膚表面状態測

定装置を使用し、培養液に対する皮膚表面の電位差（mV）を測定した。測定に用いた皮膚サンプルは、ヘアレスマウスの皮膚切片を用い、（1）皮膚表面をセロテープでバリアー破壊（テープストリッピング）させた皮膚、（2）テープストリッピングをした後、刺激阻害剤として働くことが公知のカルシウムチャネルブロッカーであるベラパミルを培養液に添加した場合の皮膚、の二種類を使用した。結果を図 3 に示す。

【0051】図 3 において、横軸はテープストリッピングしてからの経過時間を示す。なお、経過時間ゼロの測定値はテープストリッピングする前の皮膚の値である。また、ベラパミルを培養液に添加した場合は、添加した時をゼロとし、ゼロ時点の測定値はテープストリッピングする前の皮膚の値を示してある。縦軸は電位差を示す。測定値は 5 回測定した平均で表し、そのばらつきを同時に示してある。

【0052】図 3 に示されたように、テープストリッピングでの刺激によって、それ以前に比べ急激な電位差の変化を示すことが分かる。これに対して、テープストリッピング後にベラパミルを培養液に添加した場合電位差変化を抑制する。このことから、ベラパミルがテープストリッピング刺激を抑制した結果、電位差変化を減少させることが分かる。

【0053】（実施例 3）図 1 に示した皮膚表面状態測定装置を使用し、培養液に対する皮膚表面の電位差（mV）を測定した。測定に用いた皮膚サンプルは、ヘアレスマウスの皮膚切片を用い、（1）皮膚表面をセロテープでバリアー破壊（テープストリッピング）させた皮膚、（2）テープストリッピングをした後、カリウムチャネルブロッカーであり、刺激阻害効果を示すことが分かっているアミノピリジン（aminopyridine）（AP）を培養液に添加した場合の皮膚、（3）テープストリッピングをした後、ナトリウムチャネルブロッカーであり、刺激阻害効果を示さないことが分かっているテトロドトキシン（tetrodotoxin）（TTX）を培養液に添加した場合の皮膚、の三種類を使用した。結果を図 4 に示す。

【0054】図 4 において、横軸はテープストリッピングしてからの経過時間を示す。なお、経過時間ゼロの測定値はテープストリッピングする前の皮膚の値である。また、AP あるいは TTX を培養液に添付した場合は、添付した時をゼロとし、ゼロ時点の測定値はテープストリッピングする前の皮膚の値を示してある。縦軸は電位差を示す。測定値は 5 回測定した平均で表し、そのばらつきを同時に示してある。

【0055】図 4 に示されたように、テープストリッピングでの刺激によって、それ以前に比べ急激な電位差の変化を示すことが分かる。これに対して、テープストリッピング後に、AP を培養液に添加した場合電位差変化を抑制する。また、TTX を培養液に添加した場合は電

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位差変化を抑制しない。これらのことから、APがテープストリッピング刺激を抑制した結果電位差変化を減少させ、TTXは抑制しないので電位差変化を減少させないことが分かる。

【0056】（実施例4）図1に示した皮膚表面状態測定装置を使用し、培養液に対する皮膚表面の電位差（mV）を測定した。測定に用いた皮膚サンプルは、ヘアレスマウスの皮膚切片を用い、（1）皮膚表面に何も処置を施さないもの、（2）刺激物質であるカプサイシンを皮膚表面に塗布したもの、（3）カプサイシンを皮膚表面に塗布した後に、刺激の防御物質として知られているカプサゼピンを塗布したもの、の三種類を使用した。結果を図5に示す。

【0057】図5において、横軸はカプサイシンを塗布してから経過時間を示す。また、カプサゼピンを塗布した場合は、それを塗布した時点をゼロとなるようにされる。縦軸は電位差を示す。測定値は5回測定した平均で表し、そのばらつきを同時に示してある。

【0058】図5に示されたように、皮膚表面に何も処置を施さないもの（コントロール）に対して、カプサイシンを皮膚表面に塗布すると、電位差の急激な変化が見られ、塗布後0.5時間に電位差の変化が最大となっている。一方、カプサイシンを皮膚表面に塗布した後、カプサゼピンを塗布すると、電位差の変化の幅が減少し、塗布したカプサゼピンがカプサイシンによる皮膚表面の刺激を抑制した結果、電位差変化が減少していることが分かる。

【0059】以上の実施例の他、刺激物質として、ヒスタミン、ドデシルベンゼンサルフェート（界面活性剤）、乳酸（有機酸）を用いた場合も同様の結果が得られた。

【0060】

【発明の効果】以上説明した通り、本発明による皮膚表面状態測定装置によれば、培養液の液面に表面を上にして皮膚切片を浮かべた皮膚モデルを作成し、その皮膚表面と培養液との電位差を簡便に測定することができる。また、刺激要因を皮膚表面あるいは培養液に付与することにより、皮膚表面と培養液との電位差が増大し、皮膚

に与える刺激度を確認できる。さらに、刺激防御物質を皮膚に刺激要因を与える前後に、塗布したり、培養液に添加したりすることにより電位差の変化の度合いが減少し、皮膚刺激に対する抑制物質を探索することができる。

【図面の簡単な説明】

【図1】本発明の実施の形態にかかる皮膚表面状態測定装置の例を示す図である。

10 【図2】本発明による皮膚表面状態測定装置を使用して培養液に対する皮膚表面電位差を測定した結果の例を示すグラフである。

【図3】本発明による皮膚表面状態測定装置を使用して培養液に対する皮膚表面電位差を測定した結果の他の例を示すグラフである。

【図4】本発明による皮膚表面状態測定装置を使用して培養液に対する皮膚表面電位差を測定した結果の他の例を示すグラフである。

20 【図5】本発明による皮膚表面状態測定装置を使用して培養液に対する皮膚表面電位差を測定した結果の他の例を示すグラフである。

【符号の説明】

1…培養液

2…皮膚切片

3…容器

4…標準電極部

5…測定電極部

6…電位差検出部

7、8…塩橋

9…ろ紙

30 41、51…外チューブ

42、52…内チューブ

43、53…液絡

44、45、54、55…内部液

46…標準電極

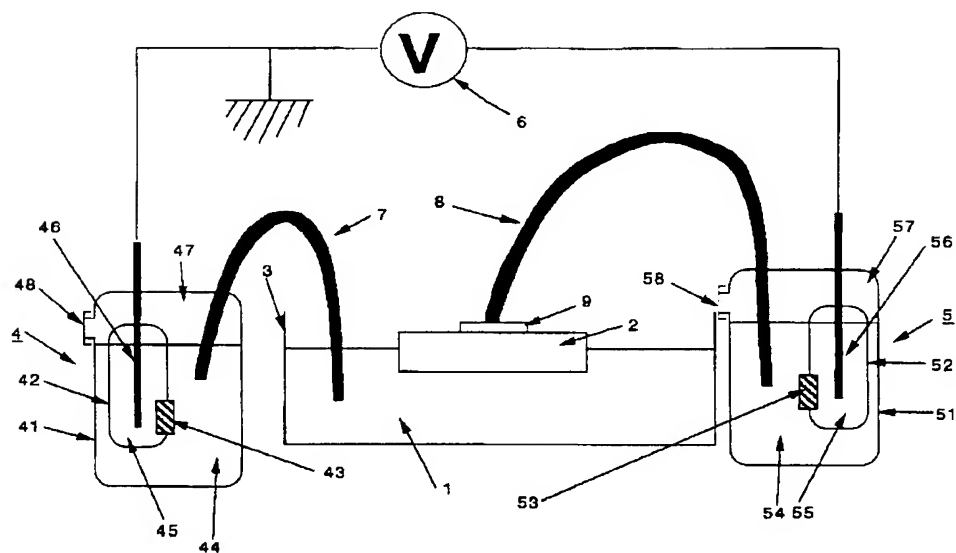
47、57…電極ホルダ

48、58…液補充口

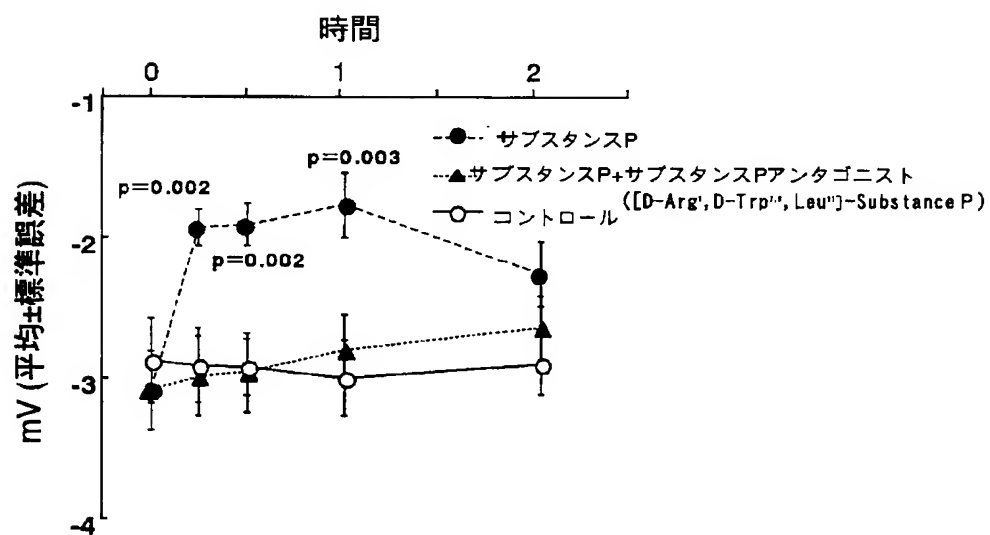
56…測定電極



【図1】

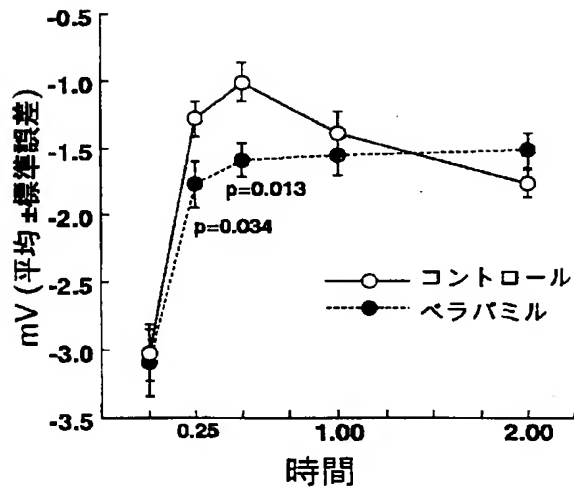


【図2】

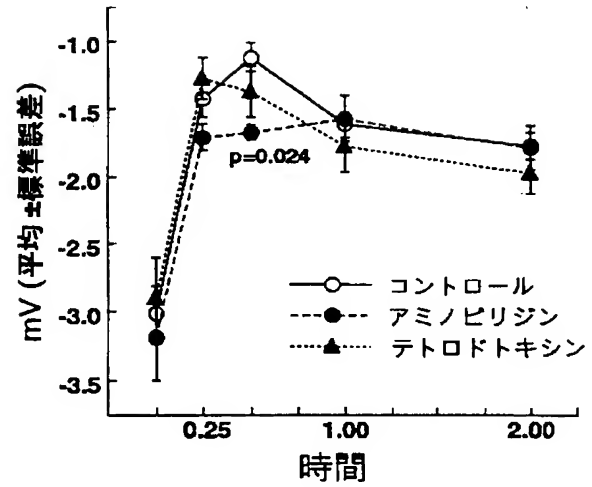




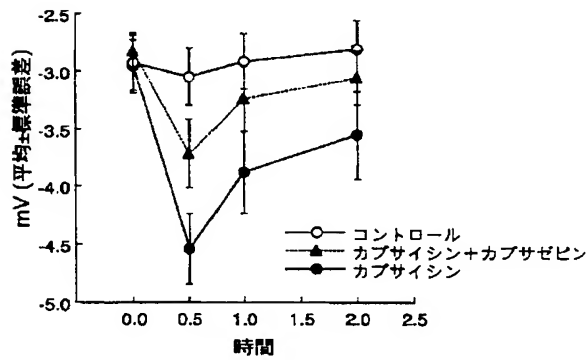
【図3】



【図4】



【図5】



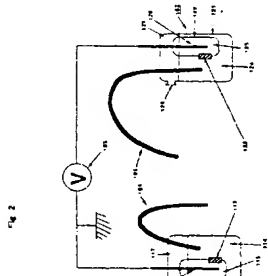
フロントページの続き

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(11) EP 1 314 979 A1

From the foregoing, the potential condition of the skin surface can be measured simply, particularly without imparting pain to a human, enabling for digitalization of the condition of stimulation to the skin surface and assessment and development of a drug for warding off stimulation.



[0012] Furthermore, the present invention is a method of floating a skin section, with its surface upward, on the surface of the culture solution accommodated in a container, and upon measuring the potential difference be-

about the human skin surface condition is useful in the field of dermatology. In the past, upon measuring the potential condition of the human skin surface, the dermis layer and the skin which has been vigorously rubbed with a file have been used as a standard. The latter case was defective in that the potential of a standard point was changes in time, and in both cases, a severe disorder was imparted to the skin due to opening in the skin a

tween the skin surface of this skin section and said culture solution, measuring the potential difference by adding a stimulating substance to the culture solution to determine a difference between the measured potential and the potential difference when no stimulating substance is added, in order to measure the degree of skin stimulation.

[0013] The stimulating substance most typically used upon measuring said degree of skin stimulation is a substance which generates an itch.

[0014] Moreover, the present invention is a method of floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container, and upon measuring the potential difference between the skin surface of this skin section and said culture solution, measuring the potential difference by coating the skin surface with a drug, either before or after the stimulation is imparted to the skin surface, to determine a difference between the measured potential difference and the potential difference when the skin surface is coated with no drug, in order to measure the degree of inhibition of the skin stimulation by a drug.

[0015] In addition, the present invention is a method of floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container, and upon measuring the potential difference between the skin surface of this skin section and said culture solution, measuring the potential difference by adding a drug to the culture solution, either before or after imparting stimulation to the skin surface, to determine a difference between the measured potential difference and the potential difference when no drug is added, in order to measure the degree of inhibition of skin stimulation by a drug.

[0016] In the aforementioned measurement of the degree of inhibition of skin stimulation, the stimulation may be caused by a stimulating substance or physical stimulation.

[0017] In addition, the present invention is a method of floating a skin section, with its surface upward, on a culture solution accommodated in a container, to which a stimulating substance is added, and upon measuring the potential difference between the skin surface of this skin section and said culture solution, measuring the potential difference by coating the skin surface with a drug, either before or after a stimulating substance is added to the culture solution, to determine a difference between the measured potential difference and the potential difference when the skin surface is coated with no drug, in order to measure the degree of inhibition of skin stimulation by a drug.

[0018] Furthermore, the present invention is a method of floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container, to which a stimulating substance is added, and upon measuring the potential difference between the skin surface of this skin section and said culture solution, measuring the potential difference by adding a drug to

the culture solution, either before or after a stimulating substance is added to the culture solution, to determine a difference between the measured potential and the potential when no stimulating substance is added, in order to determine the degree of inhibition of skin stimulation by a drug.

[0019] The stimulating substance most typically used upon measuring said degree of inhibition of skin stimulation is a substance which generates an itch.

[0020] Furthermore, the present invention is a skin surface condition measuring apparatus comprising a culture solution, a container accommodating a skin section floating, with its surface upward, on the surface of the culture solution, a standard electrode part communicated to said culture solution, a measuring electrode part communicated to the skin surface of said skin section, and a potential difference detecting part for detecting the potential difference between said standard electrode part and said measuring electrode part.

[0021] It is preferable that said standard electrode part used in the skin surface condition measuring apparatus using a culture solution is provided with an inner tube accommodating an internal solution immersed with a standard electrode, and an outer tube accommodating an internal solution which accommodates this inner tube to form a double tube, and that a liquid junction is attached to the wall of the inner tube.

[0022] In this case, the communication between the culture solution and the standard electrode part is made by a salt bridge connected between the culture solution and the internal solution of the outer tube.

[0023] In addition, said measuring electrode part used in the skin surface condition measuring apparatus may comprise an inner tube accommodating an internal solution immersed with a measuring electrode, and an outer tube accommodating an internal solution which accommodates this inner tube to form a double tube, and a liquid junction attached to the wall of the inner tube.

[0024] In this case, the communication between the skin surface of the skin section and the measuring electrode part is made by a salt bridge connected between the skin surface of the skin section and the internal solution in the outer tube.

[0025] Said skin surface condition in the apparatus of the present invention may be the potential condition of the skin surface.

[0026] Note that it is preferable that the culture solutions used in each of the aforementioned methods and each apparatus all contain physiological saline or amino acids.

[0027] According to the methods and apparatus for measuring the skin surface condition using a culture solution, the condition of the skin surface may be measured simply by measuring the potential difference between the skin surface of the skin section and the culture solution on which a skin section is floated using a suitable apparatus. Furthermore, by measuring the change in the said potential difference by imparting a stimulating

factor to the skin surface or the culture solution, stimulation may be expressed objectively without directly employing a human yet exceeding human subjectivity. In addition, the present invention may be utilized in investigating a drug which is effective to skin stimulation due to a change in the aforementioned potential difference caused by adding a drug, and which can produce a considerable effect on the investigation of new drugs, and is very useful.

[0028] Furthermore, the present invention is a method for measuring the skin surface condition, characterized by using a sublingual potential as a standard potential upon measuring of the potential condition of the skin surface.

[0029] In order to practice this measuring method, there can be adopted a method for measuring the skin surface condition from a potential difference between a standard electrode part communicating with a sublingual part and a measuring electrode part communicating with the skin surface.

[0030] Furthermore, the present invention is the skin surface condition measuring apparatus, which is provided with a standard electrode part communicating with a sublingual part, a measuring electrode part communicating with the skin surface, and a potential difference detecting part for detecting a potential difference between the standard electrode part and the measuring electrode part.

[0031] It is preferable that a standard electrode part used in this skin surface condition measuring apparatus using a sublingual potential as a standard potential is provided with an inner tube accommodating an internal solution with a standard electrode immersed therein, and an outer tube accommodating an internal solution which accommodates this inner tube to form a double tube, and a liquid junction is attached to the wall of the inner tube.

[0032] In this case, communication between the sublingual part and the standard electrode part is made by a salt bridge connecting the sublingual part and the internal solution in the outer tube of the standard electrode part.

[0033] In addition, the measuring electrode part used in the skin surface condition measuring apparatus using a sublingual potential as a standard potential may be provided with an inner tube accommodating an internal solution with a measuring electrode part immersed therein, and an outer tube accommodating an internal solution which accommodates this inner tube to form a double tube, and a liquid junction attached to the wall of the inner tube.

[0034] In this case, communication between the skin surface and the measuring electrode part is made by a salt bridge connecting the skin surface and the internal solution in the outer tube of the measuring electrode part.

[0035] The skin surface condition using said sublingual potential of the present invention as a standard po-

tential may be a potential condition of the skin surface. [0036] According to the skin surface condition measuring method and apparatus using a sublingual potential as a standard potential, by using a suitable apparatus to measure the potential difference between the skin surface and the sublingual part, the skin surface condition can be measured simply without causing pain to a human.

[0037] Note that, in the present invention, the concept of the skin surface condition includes, in addition to the potential condition of skin surface, various changes in condition occurring on skin surface, for example, the change in condition and the degree thereof due to stimulation, inhibition of stimulation and the like.

#### Brief Description of the Drawings

[0038]

Fig. 1 is a diagram showing an embodiment of the skin surface condition measuring apparatus using a culture solution according to the present invention.

Fig. 2 is a diagram showing an embodiment of the skin surface condition measuring apparatus using a sublingual potential as a standard potential according to the present invention.

Fig. 3 is a graph showing an example of the results obtained by measuring a skin surface potential difference relative to a culture solution employing the skin surface condition measuring apparatus using a culture solution according to the present invention.

Fig. 4 is a graph showing another example of the results obtained by measuring a skin surface potential difference relative to a culture solution employing the skin surface condition measuring apparatus using a culture solution according to the present invention.

Fig. 5 is a graph showing another example of the results obtained by measuring a skin surface potential difference relative to a culture solution employing the skin surface condition measuring apparatus using a culture solution according to the present invention.

Fig. 6 is a graph showing another example of the results obtained by measuring a skin surface potential difference relative to a culture solution employing the skin surface condition measuring apparatus using a culture solution according to the present invention.

Fig. 7 is a graph showing an example employing the skin surface condition measuring apparatus using a sublingual potential as a standard potential according to the present invention.

Fig. 8 is a graph showing the correlation between a skin surface potential employing the skin surface condition measuring apparatus using a sublingual

potential as a standard potential according to the present invention, and a skin surface potential according to the conventional method.

#### Best Mode for Carrying Out the Invention

[0039] Embodiments of the present invention will be described in detail below. First, a method and apparatus for measuring the skin surface condition using a culture solution will be explained.

[0040] This method of the present invention includes several methods. In a first method, a skin section is floated, with its surface upward, on the surface of a culture solution accommodated in a container, and a potential difference (mV) between the skin surface of this skin section and the culture solution is measured, thereby measuring the surface condition of skin.

[0041] In a second method, upon measuring the skin surface condition of the first method, stimulation is imparted further to the skin model comprising a skin section and a culture solution to measure aforementioned potential difference. This potential difference changes as stimulation is imparted. This change means that a stream of ions has occurred in the epidermis. The direction of a stream of ions differs depending on stimulation. When there is a change compared to the value of untreated skin, it is interpreted that stimulation producing mobility of ions is imparted to the epidermis. The degree of skin stimulation is measured from the changed value. Upon imparting stimulation, the place of stimulation may be on skin surface or culture solution.

In the case of skin surface, examples of methods for imparting stimulation include coating skin with a stimulating substance and physical stimulation. Also, in the case of culture solution, the method used is addition of stimulating substances. Examples of these stimulating substances include neuropeptide, histamine, surfactant, organic acid, and capsaicin. Among them, neuropeptide includes substance P. In the present invention, it is particularly useful that the stimulating substance generates an itch. In addition, examples of said physical stimulation include barrier destruction by a cellophane adhesive tape and the like.

[0042] A third method is a method for measuring the degree of inhibition by addition of a drug to the stimulated skin in the second method. In the present invention, after the aforementioned skin model is stimulated, a drug is added to measure the aforementioned potential difference, and the degree of inhibition of skin stimulation by a drug is measured from the value of potential difference which changes by addition. The present method can take various methods depending on a method of drug addition. More specifically,

(1) A skin section, the surface of which has been stimulated, is floated with its surface upward, on a culture solution, a potential difference is measured by coating skin surface with a drug, either before or after imparting skin stimulation, and the degree of inhibition of

skin stimulation by a drug is determined from a value of potential difference changed by drug coating. (2) In place of coating the skin surface in (1) with a drug, a drug is added to a culture solution to measure a potential difference, and the degree of skin stimulation by a drug is measured from the value of the potential difference, which changes by drug addition to the culture solution.

(3) A skin section is floated, on a culture solution with a stimulating substance added thereto, with the surface upward, a potential difference is measured by coating a drug on the skin surface either before or after addition of a stimulating substance, and the degree of inhibition of skin stimulation by a drug is determined from a value of the potential difference changed by drug coating. (4) In place of coating the skin surface in (3) with a drug, a drug is added to a culture solution to measure a potential difference, and the degree of inhibition of skin stimulation by a drug is measured from the value of the potential difference, which changes by drug addition to the culture solution.

[0043] In addition, the aforementioned coating of a drug may be performed either before or after imparting stimulation. When the drug is coated after imparting stimulation, the effect of alleviating the stimulation can be measured, and when the drug is coated before imparting stimulation, the effect of warding off the stimulation beforehand can be measured.

[0044] The culture solution used in the measuring methods 1 through 3 may be physiological saline and an aqueous solution containing amino acids and the like. In addition, the skin section used in the measuring method may be a cultured skin, a human skin obtained on plastic surgery and the like, and a skin derived from an animal such as a hairless mouse.

[0045] When a human skin is stimulated, a symptom appears on the skin, and at the same time, an itch, a pain, a smarting pain and the like, which are expressed by human senses, are produced in many cases. Also in many cases, no symptom appears but only the sense is produced. The aforementioned 1 through 3 methods of the present invention, enable such senses, which are subjective, to be expressed by numerical values, which are objective. As a result, for example, these methods can be utilized in searching for a drug which either alleviates or inhibits stimulation without employing a human.

[0046] Next, an apparatus which is preferably used in the method using a culture solution of the present invention will be explained with reference to the drawings.

[0047] Fig. 1 shows an embodiment of the skin surface condition measuring apparatus using a culture solution in the present invention. As shown in Fig. 1, the skin surface condition measuring apparatus has a culture solution 1, a container 3 accommodating a skin section 2 floated, with its surface upward, on the surface of the culture solution, a standard electrode part 4 communicating the culture solution 1, a measuring electrode part 5 communicating to the skin surface of the skin section

tion 2, and a potential difference detecting part 6 for detecting a potential difference between the standard electrode part 4 and the measuring electrode part 5.

[0048] The standard electrode part 4 has a double outer tube 41 and inner tube 42 which are made of a plastic, and a tube wall of the inner tube 42 has a liquid junction 43 composed of a porous ceramic having the function of not permeating a solution but permeating electrons or ions. The interiors of the double outer tube 41 and inner tube 42 are filled with internal solutions 44, 45 comprising a KC1 solution having the predetermined concentration, respectively, and a standard electrode 46 comprising silver-silver chloride is immersed in the internal solution 45 of the inner tube 42. This standard electrode 46 is attached to an electrode holder 47 which also functions as a cap and closes upper ends of the double outer tube 41 and inner tube 42, and in the condition where the electrode is suspended from there via a conductor, the electrode is immersed in the internal solution 45 in the inner tube 42. A solution supplementing port 48 for supplementing the internal solution 44 in the outer tube 41 is provided on a circumferential plane near an upper end of the outer tube 41.

[0049] Note that, in the standard electrode part 4 in Fig. 1, the double outer tube 41 and inner tube 42 may all be made of glass. In addition, the liquid junction 43 used may be a known liquid junction such as a solution having a pseudo-crystal structure such as gelatin and agar, and may include a liquid junction used as a salt bridge such as a porous Bicolli glass. Alternatively, the standard electrode 46 used may be an electrode comprising of calomel. In addition, the internal solution 45 used may be other than a KC1 solution, a solution of chloride such as sodium chloride, lithium chloride and the like. The internal solution 44 in the outer tube 41 used may be, in addition to chloride used in the internal solution 45 in the inner tube 42, physiological saline.

[0050] The measuring electrode part 5 has the similar structure as that of the standard electrode part 4. More specifically, the measuring electrode part 5 has a double outer tube 51 and an inner tube 52 made of a plastic, and the tube wall of an inner tube 52 has a liquid junction 53 composed of a porous ceramic. The interiors of the double outer tube 51 and the inner tube 52 are filled with internal solutions 54 and 55, respectively, comprising a KC1 solution of a predetermined concentration, and a measuring electrode 56 composed of silver-silver chloride is immersed in the internal solution 55 of the inner tube 52. This measuring electrode 56 is attached to an electrode holder 57 which also functions as a cap to close upper ends of the double outer tube 51 and the inner tube 52, and is immersed in the internal solution 55 in the inner tube 52 in a state of being suspended from the electrode holder via a conductor. A solution supplementing port 58 for supplementing the internal solution 54 in the outer tube 51 is provided on a circumferential plane near an upper end of the outer tube 51.

[0051] Note that, the measuring electrode part 5 in

Fig. 1 may also take an embodiment which is similar to that of the standard electrode part 4. More specifically, the double outer tube 51 and the inner tube 52 used may be all made of glass. In addition, the liquid junction 53 used may be the porous Bicolli glass. In addition, the aforementioned measuring electrode 56 used may comprise of calomel. In addition, the internal solution 55 used may be other than a KC1 solution, a solution of chloride such as sodium chloride and lithium chloride. The internal solution 55 used in the outer tube 51 may be other than chloride used in said internal solution 55, physiological saline.

[0052] Furthermore, the standard electrode part 4 in Fig. 1 is communicated to the culture solution 1, which comprises of DMEM (Dulbecco's modified Eagle's medium), via a salt bridge 7, in which a flexible plastic tube is immersed in the internal solution 44 in its outer tube 41 is filled with agarose containing physiological saline. This DMEM is a known culture solution containing amino acids and is commercially available. Other known culture solutions such as those containing physiological saline may also be used. In addition, the salt bridge 7 may be a known one, and for example, rubber tubes such as silicone and the like may be used.

[0053] In addition, the measuring electrode part 5 is communicated to the skin surface of the skin section 2 through a filter 9 placed on the skin surface via a salt bridge 8 in which a flexible plastic tube immersed in an internal solution 54 in the outer tube 51 is filled with agarose containing physiological saline. This filter 9 achieves uniformity in the contact of the salt bridge 8 with the skin surface of skin section 2. In addition, the salt bridge 8 may be a known one, and may for example use rubber tubes such as silicone and the like.

[0054] The standard electrode 46 of the aforementioned standard electrode part 4 and the measuring electrode 56 of the measuring electrode 5 are connected to a potential difference detecting part 6 to measure a potential difference thereof.

[0055] Note that, the standard electrode part used in the skin surface condition measuring apparatus may be provided with a liquid junction on the tube wall of the outer tube 41 of the standard electrode part 4 in Fig. 1. The outer tube may be provided with a solution supplementing port, which, like in Fig. 1, supplements an internal solution.

[0056] This standard electrode part has a function similar to that of the standard electrode part 4 of the skin surface condition measuring apparatus in Fig. 1, and it can be used in place of the standard electrode part 4 in Fig. 1, provided that communication is performed by immersion of this standard electrode part in the culture solution and via a liquid junction provided on an outer tube wall. Note that, embodiments of this standard electrode part are the same as those in Fig. 1.

[0057] Furthermore, the standard electrode part in the skin surface condition measuring apparatus may comprise of only the inner tube 42 of the standard electrode

part 4 in Fig. 1, namely, the inner tube 42, the liquid junction 43, the internal solution 45 and the standard electrode 46.

[0058] This standard electrode part also has a function similar to that of a standard electrode part 4 of the skin surface condition measuring apparatus in Fig. 1, and it can be used in place of the standard electrode part 4 in Fig. 1, provided that, communication with a culture solution is performed by immersion of this standard electrode part into the culture solution and via a liquid junction provided on the tube wall. In addition, embodiments in this standard electrode part are the same as in Fig. 1.

[0059] Next, a method and apparatus for measuring the skin surface condition using a sublingual potential as a standard potential will be explained.

[0060] In this method of the present invention, a sublingual area is used as a standard to measure the potential difference (mV) between the skin surface and the sublingual area. As a result, the skin surface condition is measured.

[0061] Next, an apparatus which is suitably used in the aforementioned method using a sublingual potential as a standard potential of the present invention will be explained with reference to the drawings.

[0062] Fig. 2 shows an example of the skin surface condition (potential condition of the skin surface) using a sublingual potential as a standard potential in the present invention. As shown in Fig. 2, the skin surface condition measuring apparatus has a standard electrode part 101 communicating to a sublingual area (not shown), a measuring electrode part 102 communicating to the skin surface (not shown), and a potential difference detecting part 103 for detecting a potential difference between the standard electrode part 101 and the measuring electrode part 102.

[0063] The standard electrode part 101 has double outer tube 111 and inner tube 112 made of a plastic, and a tube wall of the inner tube 112 has a liquid junction

113 comprising a porous ceramic having the function of not permeating a solution but permeating electrons or ions. The interiors of the double outer tube 111 and inner tube 112 are filled with internal solutions 114, 115 comprising a KC1 solution having the prescribed concentration, respectively, and a standard electrode 116 composed of silver-silver chloride is immersed in the internal solution 115 of the inner tube 112.

This standard electrode 116 is attached to an electrode holder 117 which also functions as a cap and closes upper ends of the double outer tube 111 and inner tube 112, and in the condition where the electrode is suspended from there via a conductor, the electrode is immersed in the internal solution 115 in the inner tube 112. A solution supplementing port 118 for supplementing the internal solution 114 in the outer tube 111 is provided on a circumferential plane near an upper end of the outer tube 111.

[0064] In addition, in the standard electrode part 101 in Fig. 2, the double outer tube 111 and inner tube 112

all made of glass may be used. In addition, as the liquid junction 113, the known liquid junctions such as solutions having the pseudo-crystal structure such as gelatin, agar and the like and liquid junctions including those used as a salt bridge such as porous Bicol glass and the like may be used. Alternatively, as the standard electrode 116, an electrode comprising calomel may be used. In addition, as the internal solution 115, a solution of chloride such as sodium chloride, lithium chloride and the like can be used in addition to a KC1 solution. As the internal solution 114 in the outer tube 111, physiological saline may be used in addition to chloride used in the internal solution 115 in the inner tube 112.

[0065] The measuring electrode part 102 has the similar structure to that of the standard electrode part 101. That is, the measuring electrode part 102 has double outer tube 121 and inner tube 122 made of a plastic, and a tube wall of the inner tube 122 has a liquid junction 123 composed of a porous ceramic. The interiors of the double outer tube 121 and inner tube 122 are filled with internal solutions 124, 125 comprising a KC1 solution having the prescribed concentration, respectively, and a measuring electrode 126 composed of silver-silver chloride is immersed in the internal solution 125 of the inner tube 122. This measuring electrode 126 is attached to an electrode holder 127 which also functions as a cap and closes upper ends of the double outer tube 121 and inner tube 122, and in the condition where the electrode is suspended from there via a conductor, the electrode is immersed in the internal solution 125 in the inner tube 122. A solution supplementing port 128 for supplementing the internal solution 124 in the outer tube 121 is provided on a circumferential plane near an upper end of the outer tube 121.

[0066] Note that the measuring electrode part 102 in Fig. 2 can also take the similar embodiment to that of the standard electrode part 101. That is, the double outer tube 121 and inner tube 122 all made of glass may be used. In addition, porous Bicol glass may be used in the liquid junction 123. In addition, as the measuring electrode 126, an electrode comprising calomel may be used. In addition, as the internal solution 125, a solution of chloride such as sodium chloride, lithium chloride and the like can be used in addition to a KC1 solution. As the internal solution 124 in the outer tube 121, physiological saline may be used in addition to chloride used in the internal solution 125.

[0067] Furthermore, the standard electrode part 101 in Fig. 2 is communicated to a sublingual area via a salt bridge 104 immersed in the internal solution 114 in its outer tube 111. The salt bridge 104 may be the known one, and for example, flexible plastic tubes, rubber tubes such as silicone and the like, and the like which are filled with agarose containing physiological saline are used.

[0068] In addition, the measuring electrode part 102 is communicated to the skin surface via a salt bridge 105 immersed in the inner solution 124 in its outer tube

121. The salt bridge 105 may be the known one, and for example, flexible plastic tubes, rubber tubes such as silicone and the like, and the like which are filled with agarose containing physiological saline are used.

[0069] The standard electrode 116 of the standard measuring electrode part 102 and the measuring electrode 126 of the measuring electrode part 102 are connected to the potential difference detecting part 103, respectively.

[0070] In order to measure the skin surface condition (potential condition) using the aforementioned apparatus, one end of a salt bridge, the other end of which is communicated to a standard electrode part, is placed in mouth of a subject to be measured, and is fixed sublingual. Upon this, one end of a salt bridge is not directly placed in mouth, but a tube such as a plastic, a silicon rubber and the like containing therein a cotton or the like containing exchangeable physiological saline is connected to an end of a salt bridge, and the tube may be placed in mouth and fixed sublingual. By doing so, since the tube can be exchanged every subject to be measured, there is no unclear feeling, and in particular, the tube is effective upon use at shop fronts and clinical places.

On the other hand, after one end of a salt bridge, the other end of which is communicated to a measuring electrode part, is contacted with and fixed on the skin surface of a subject to be measured, a potential difference between the standard electrode part and the measuring electrode part is measured with a potential difference detecting part. Note that it is preferable that communication between the salt bridge and the skin surface is through a filter in order to contact both uniformly.

[0071] According to the method for measuring the skin surface condition using a sublingual potential as a standard potential, a number of information which are useful for dermatology can be obtained. For example, physical stimulation such as barrier destruction with a cellophane adhesive tape and the like is given to the skin surface, or chemical stimulation is given to the skin surface by coating a stimulating substance such as neuropeptide, histamine, surfactant, organic acid, capsaicin and the like, a potential of the skin surface having subjective sense such as itching and the like is measured, and the skin surface condition can be measured by expressing a change in the skin surface condition as an objective numerical value from comparison of potential differences before and after stimulation.

[0072] Furthermore, a potential difference is measured by coating adjuvant on the stimulated skin surface, and the degree of inhibition of skin stimulation by a drug may be measured from a value of a potential difference changed by drug coating.

[0073] Then, more specific Examples of the present invention will be explained.

(Example 1)

[0074] A potential difference (mV) of the skin surface relative to a culture solution was measured using the

skin surface condition measuring apparatus shown in Fig. 1. The skin samples used in measurement are three kinds of skin sections of a hairless mouse, which are (1) a sample in which the skin surface is not treated, (2) a sample in which substance P (SP) of a stimulating substance is added to a culture solution, and (3) a sample in which SP antagonist (ID-AngI, D-Trp7, 9, LeuIIl -SubstanceP), which, together with SP, is known as a substance for warding off stimulation, is added to a culture solution. The results are shown in Fig. 3.

[0075] In Fig. 3, the horizontal axis indicates the elapsed time after addition of SP. In addition, when an SP antagonist is added, the time point of addition is indicated as zero. The vertical axis indicates potential differences. A measured value is expressed as an average of five measurements, and the fluctuation is indicated simultaneously.

[0076] As shown in Fig. 3, when SP is added to a culture solution, a rapid change in a potential difference is produced as compared with the non-treated skin surface (control). The state with a significant potential difference continues, reaching its peak 1 hour after the addition, until 2 hours after the addition when the potential difference returns to a value close to that of a control. On the other hand, when an SP antagonist is added to a culture solution before adding SP, no rapid and significant change in potential difference is produced, and a value almost the same as that of a control is indicated for 2 hours after the addition. From this, it can be seen that, when SP and an SP antagonist are added together, it can be seen that, an SP antagonist inhibits stimulation by SP, and inhibits a change in a potential difference.

(Example 2)

[0077] A potential difference (mV) of the skin surface relative to a culture solution was measured using the skin surface condition measuring apparatus shown in Fig. 1. The skin samples used in measurement are two kinds of skin sections of a hairless mouse, which are (1) a skin with its surface barrier destructured with a cellophane adhesive tape (tape stripping), and (2) a skin after tape stripping, with verapamil, which is a potassium channel blocker known to function as an agent for inhibiting stimulation, added to a culture solution afterwards. The results are shown in Fig. 4.

[0078] In Fig. 4, the horizontal axis indicates the elapsed time after tape stripping. In addition, the measured value at time zero is the value of skin before applying tape stripping. In addition, when verapamil is added to a culture solution, the time of addition is indicated as zero, and the measured value at a time point zero indicates the value of skin before applying tape stripping. The vertical axis indicates potential differences. A measured value is expressed as an average of five measurements, and the fluctuation is indicated simultaneously.

[0079] As shown in Fig. 4, it can be seen that the stim-

ulation by tape stripping produces a rapid change in potential difference as compared with before the stimulation. To the contrary, adding verapamil to a culture solution after tape stripping inhibits the change in potential difference. From this, it can be seen that, as a result of tape stripping inhibiting tape stripping stimulation, the change in potential difference is reduced.

(Example 3)

[0080] A potential difference (mV) of the skin surface relative to a culture solution was measured using the skin surface condition measuring apparatus shown in Fig. 1. The skin samples used in measurement are three kinds of skin sections of a hairless mouse, which are (1) a skin with its surface barrier-destroyed with a cellophane adhesive tape (tape stripping), (2) a skin after tape stripping, with aminopyridine (AP), which is a potassium channel blocker and is known to show the stimulation inhibiting effect, added to a culture solution afterwards, and (3) a skin after tape stripping, with tetrodotoxin (TTX), which is a sodium channel blocker known not to show the stimulation inhibiting effect, added to a culture solution. The results are shown in Fig. 5.

[0081] In Fig. 5, the horizontal axis indicates the elapsed time after tape stripping. Note that the measured value at time zero is the value of skin before applying tape stripping. In addition, when AP or TTX is added to a culture solution, the time of addition is indicated as zero, and the measured value at time point zero indicates the value of skin before applying tape stripping. The vertical axis indicates potential differences. A measured value is expressed as an average of five measurements, and its fluctuation is indicated simultaneously.

[0082] As shown in Fig. 5, it can be seen that the tape stripping stimulation produces a rapid change in a potential difference as compared with before stimulation. To the contrary, adding AP to a culture solution after tape stripping inhibits the potential difference. In addition, adding TTX to a culture solution does not inhibit the change in a potential difference. From these, it can be seen that AP reduces the change in potential difference as a result of inhibiting tape stripping stimulation, and that TTX does not reduce the change in potential difference because it does not inhibit tape stripping stimulation.

(Example 4)

[0083] A potential difference (mV) of the skin surface relative to a culture solution was measured using the skin surface condition measuring apparatus shown in Fig. 1. The skin samples used in measurement are three kinds of skin sections of a hairless mouse, which are (1) a sample in which the skin surface is not treated, (2) a sample in which the skin surface is coated with capsaicin of a stimulating substance, and (3) a sample

in which capsaicin is coated on the skin surface, and capsaizepine known as a stimulation-defending substance is coated thereon afterwards. The results are shown in Fig. 6.

[0084] In Fig. 6, the horizontal axis indicates the elapsed time after coating of capsaicin. In addition, when capsaizepine is coated, the time point of coating is indicated as zero. The vertical axis indicates a potential difference. The measured value is expressed as an average of five measurements, and the fluctuation is indicated simultaneously.

[0085] As shown in Fig. 6, when capsaicin is coated on skin surface, a rapid change in potential difference reaching its maximum 0.5 hour after the coating is perceived as compared with non-treated skin surface (control). On the other hand, it can be seen that, coating skin surface with capsaicin, followed by coating of capsaizepine, the range of change in potential difference is reduced, and the change in potential difference is reduced as a result of the coated capsaizepine inhibiting the skin surface stimulation by capsaicin.

[0086] Other than the above Examples, similar results were obtained when histamine, dodecylbenzene sulfate (surfactant), or lactic acid (organic acid) was used as a stimulating substance.

(Example 5)

[0087] The potential differences of skin surface relative to a sublingual area and a scar on the skin (epidermis) were measured using the skin surface measuring apparatus shown in Fig. 2. The skin used in measurement is a skin in a state of 2 hours after skin surface was barrier-destroyed (tape stripped) with a cellophane adhesive tape. Fig. 7 shows potentials at several places of skin using each of a sublingual potential and a skin scar potential as a standard potential.

[0088] As shown in Fig. 7, a potential rises 2 hours after tape stripping. It can be seen that this rise in potential is detectable using either a sublingual area or a skin scar as a standard. In addition, letters A to G in the figure denote measurement points, and AVE denotes an average value.

[0089] Furthermore, Fig. 8 plots the results of Fig. 7 in connection with the relationship between a skin surface potential using a skin scar as a standard and a skin surface potential using a sublingual area as a standard. It can be seen that there is significant correlation between both potentials, and also when a sublingual area is used as a standard in place of the previous method using a skin scar as a standard, a skin surface potential can be stably measured.

Industrial Applicability

[0090] As explained above, according to the skin surface condition measuring apparatus of the present invention, by making a skin model, in which a skin section

is floated with its surface upward on the surface of a culture solution, a potential difference between the skin surface and the culture solution can be measured simply. Furthermore, by adding a stimulating factor to the skin surface or a culture solution, a potential difference between the skin surface and a culture solution is increased, and as a result, the degree of stimulation given to skin can be confirmed. In addition, by coating a substance for warding off stimulation on skin, or by adding it to a culture solution, either before or after impartation of a stimulating factor to skin, or by adding a substance to ward off stimulation, the degree of a change in a potential difference is decreased, and an inhibiting substance to skin stimulation can be investigated.

[0091] Furthermore, according to the skin surface condition (potential condition) measuring apparatus, which uses a sublingual potential as a standard, the skin surface condition can be measured simply without imparting pain to a human. From this result, a number of useful applications can be expected such as confirmation of the degree of stimulation to be imparted to skin, investigation of an inhibiting substance to skin stimulation from a change in a potential difference by coating a substance for warding off stimulation on skin, and the like.

## Claims

1. A method for measuring the skin surface condition, which comprises: floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container; and measuring a potential difference between the skin surface of the skin section and said culture solution.
2. The method for measuring the skin surface condition according to claim 1, wherein said culture solution is a culture solution containing physiological saline or amino acids.
3. The method for measuring the skin surface condition according to claim 1 or 2, wherein the skin surface condition is a potential condition of the skin surface.
4. A method for measuring the degree of skin stimulation, which comprises: floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container; measuring a potential difference by imparting stimulation to the skin surface upon measuring of a potential difference between the skin surface of the skin section and said culture solution; and determining the degree of inhibition of skin stimulation by the drug from a difference between the measured potential difference value and a potential difference when no drug is added.
5. The method for measuring the degree of skin stimulation according to claim 4, wherein said stimulation is made by a stimulating substance.
6. The method for measuring the degree of skin stimulation according to claim 4, wherein said stimulation is made by physical stimulation.
7. A method for measuring the degree of skin stimulation, which comprises: floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container; measuring a potential difference by adding a stimulating substance to a culture solution upon measuring a potential difference between the skin surface of the skin section and said culture solution; and determining the degree of inhibition of skin stimulation by the drug from a difference between the measured potential difference value and a potential difference when no stimulating substance is added.
8. The method for measuring the degree of skin stimulation according to claim 5 or 7, wherein the said stimulating substance is a substance which generates an itch to skin.
9. The method for measuring the degree of skin stimulation according to any one of claims 4 through 8, wherein the said culture solution is a culture solution containing physiological saline or amino acids.
10. A method for measuring the degree of inhibition of skin stimulation by a drug, which comprises: floating a stimulated skin section, with its surface upward, on the surface of a culture solution accommodated in a container; measuring a potential difference by coating a drug on the skin surface, either before or after imparting stimulation to the skin surface upon measuring a potential difference between the skin surface of the skin section and said culture solution; and determining the degree of inhibition of skin stimulation by the drug from a difference between the measured potential difference value and a potential difference when no drug is coated.
11. A method for measuring the degree of inhibition of skin stimulation by a drug, which comprises: floating a stimulated skin section, with its surface upward, on the surface of a culture solution accommodated in a container; measuring a potential difference by adding a drug to the culture solution, either before or after imparting stimulation to the skin surface upon measuring a potential difference between the skin surface of the skin section and said culture solution; and determining the degree of inhibition of skin stimulation by the drug from a difference between the measured potential difference value and a potential difference when no drug is added.

12. The method for measuring the degree of inhibition of skin stimulation according to claim 10 or 11, wherein the said stimulation is made by a stimulating substance.

13. The method for measuring the degree of inhibition of skin stimulation according to claim 10 or 11, wherein the said stimulation is made by physical stimulation.

14. A method for measuring the degree of inhibition of skin stimulation by a drug, which comprises: floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container, with a stimulating substance added thereto; measuring a potential difference by coating a drug on the skin surface either before or after adding the stimulating substance to the culture solution upon measuring a potential difference between the skin surface of the skin section and said culture solution; and determining the degree of inhibition of skin stimulation by the drug from a difference between the measured potential difference value and a potential difference when no drug is coated.

15. A method for measuring the degree of skin stimulation by a drug, which comprises: floating a skin section, with its surface upward, on the surface of a culture solution accommodated in a container, with a stimulating substance added thereto; measuring a potential difference by adding a drug to the culture solution either before or after adding the stimulating substance to the culture solution upon measuring a potential difference between the skin surface of the skin section and said culture solution; and determining the degree of inhibition of skin stimulation by the drug from a difference between the measured potential difference value and a potential difference when no stimulating substance is added.

16. The method for measuring the degree of inhibition of skin stimulation according to claim 12, 14 or 15, wherein the said stimulating substance is a substance which generates an itch to skin.

17. The method for measuring the degree of inhibition of skin stimulation according to any one of claims 10 through 16, wherein the said culture solution is a culture solution containing physiological saline or amino acids.

18. The skin surface condition measuring apparatus, which comprises: a culture solution; a container accommodating a skin section floated with its surface upward on the surface of the culture solution; a standard electrode part communicating to said culture solution; a measuring electrode part communicating

calling to the skin surface of said skin section; and a potential difference detecting part for detecting a potential difference between said standard electrode part and said measuring electrode part.

19. The skin surface condition measuring apparatus according to claim 18, wherein the said standard electrode part is provided with an inner tube accommodating an internal solution with a standard electrode immersed therein, and an outer tube accommodating an internal solution which accommodates the inner tube to form a double tube, and a liquid junction is attached to the wall of said inner tube.

20. The skin surface condition measuring apparatus according to claim 19, wherein communication between the said culture solution and the standard electrode part is made by a salt bridge connecting said culture solution and the internal solution of said outer tube.

21. The skin surface condition measuring apparatus according to any one of claims 18 through 20, wherein the said measuring electrode part is provided with an inner tube accommodating an internal solution with a measuring electrode immersed therein, and an outer tube accommodating an internal solution which accommodates the inner tube to form a double tube, and a liquid junction is attached to the wall of said inner tube.

22. The skin surface condition measuring apparatus according to claim 21, wherein communication between the skin surface of said skin section and the measuring electrode part is made by a salt bridge connecting the skin surface of said skin section and the internal solution in said outer tube.

23. The skin surface condition measuring apparatus according to any one of claims 18 through 22, wherein the said culture solution is a culture solution containing physiological saline or amino acids.

24. The skin surface condition measuring apparatus according to any one of claims 18 through 23, wherein the skin surface condition is the potential condition of the skin surface.

25. A method for measuring the skin surface condition, which comprises using a sublingual potential as a standard potential upon measuring the skin surface condition.

26. A method for measuring the skin surface condition, which comprises measuring the skin surface condition from a potential difference between a standard electrode part communicating with a sublingual area and a measuring electrode part communicating

to the skin surface.

27. The method for measuring the skin surface condition according to claim 25 or 26, wherein the skin surface condition is a potential condition of the skin surface.

28. The skin surface condition measuring apparatus, which comprises a standard electrode part communicating to a sublingual area, a measuring electrode part communicating to the skin surface, and a potential difference detecting part for detecting a potential difference between said standard electrode part and said measuring electrode part.

29. The skin surface condition measuring apparatus according to claim 28, wherein the standard electrode part is provided with an inner tube accommodating an internal solution with a standard electrode immersed therein, and an outer tube accommodating an internal solution which accommodates the inner tube to form a double tube, and a liquid junction is attached to the wall of said inner tube.

30. The skin surface condition measuring apparatus according to claim 29, wherein communication between said sublingual area and said standard electrode part is made by a salt bridge connecting the sublingual area and the internal solution in the outer tube of the standard electrode part.

31. The skin surface condition measuring apparatus according to any one of claims 28 through 30, wherein the measuring electrode part is provided with an inner tube accommodating an internal solution with a measuring electrode immersed therein, and an outer tube accommodating an internal solution which accommodates the inner tube to form a double tube, and a liquid junction is attached to the wall of said inner tube.

32. The skin surface condition measuring apparatus according to claim 31, wherein communication between said skin surface and said measuring electrode part is by a salt bridge connecting the skin surface and the internal solution in the outer tube of the measuring electrode part.

33. The skin surface condition measuring apparatus according to any one of claims 28 through 32, wherein the skin surface condition is a potential condition of the skin surface.



Fig. 1

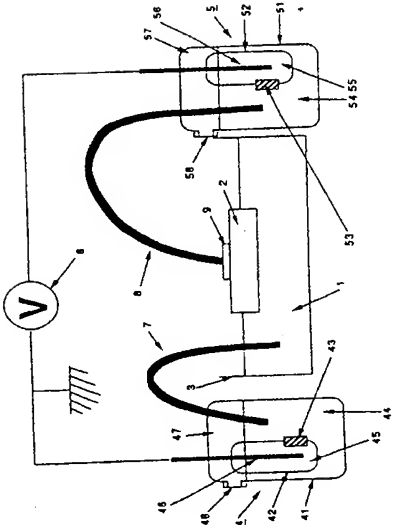
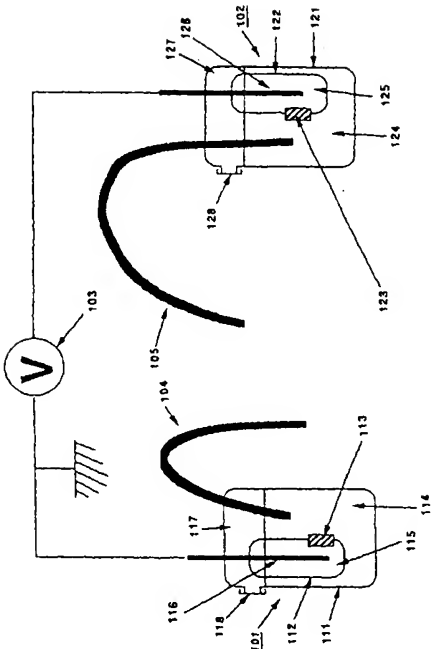


Fig. 2



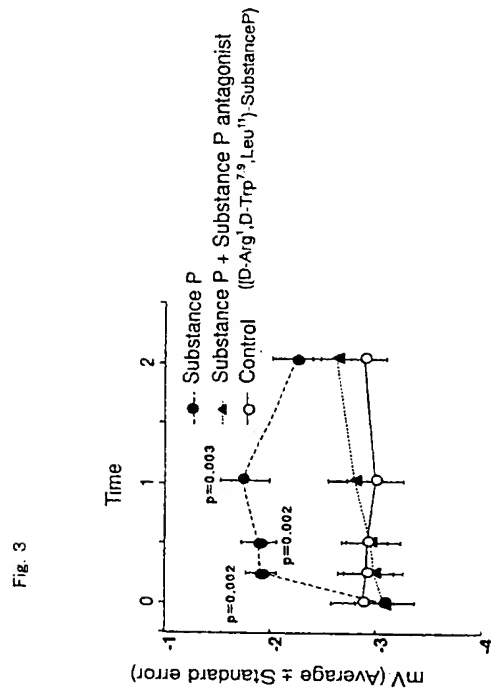


Fig. 4

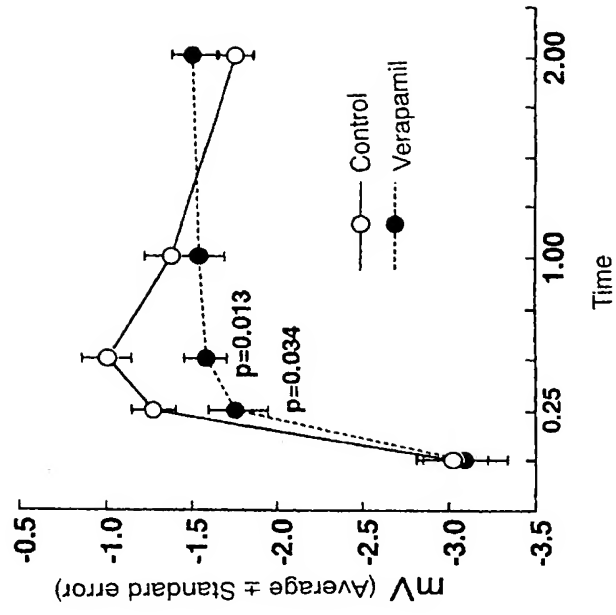


Fig. 5

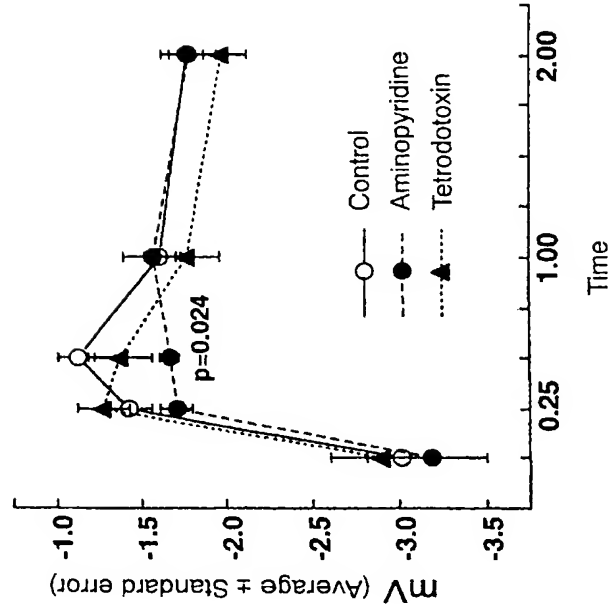


Fig. 6

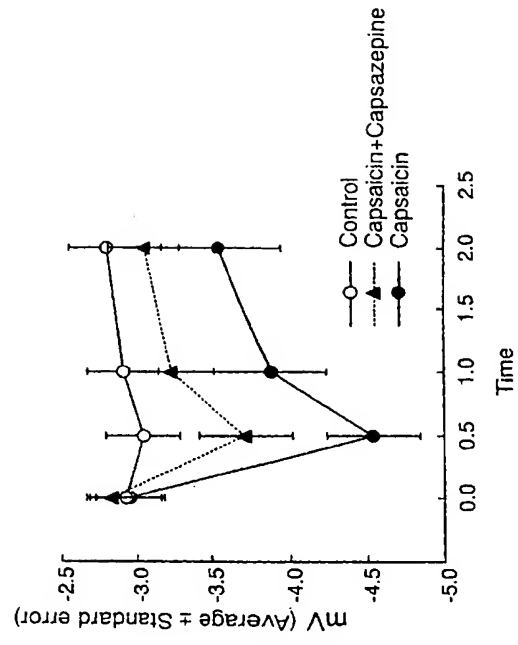


Fig. 7

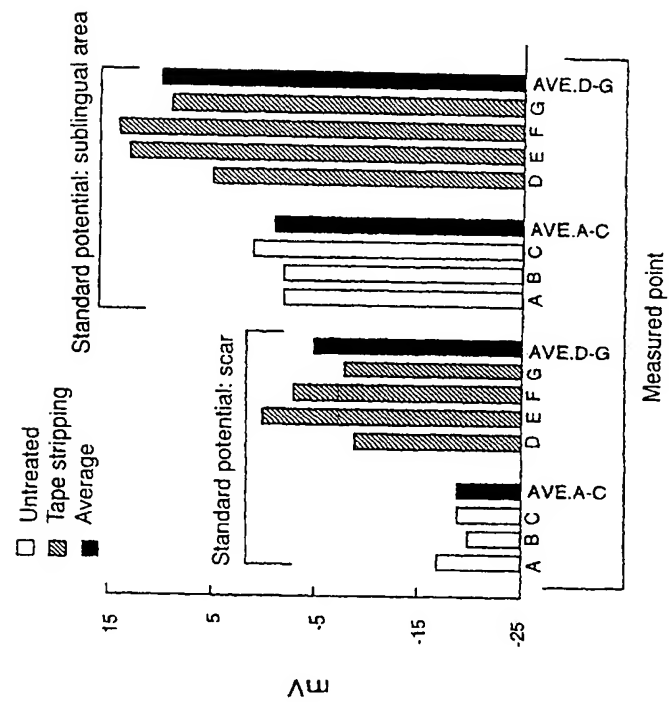
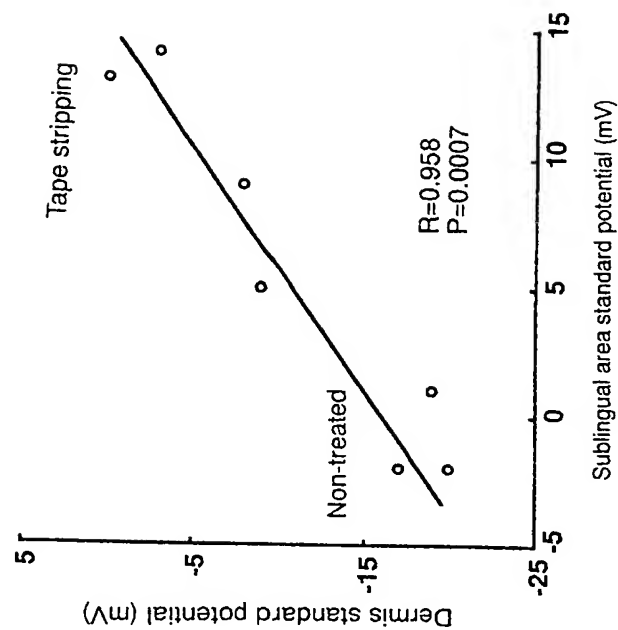


Fig. 8



## INTERNATIONAL SEARCH REPORT

International application No. PCT/JP01/07142	
A. CLASSIFICATION OF SUBJECT MATTER Int. Cl. G01N33/483, A61B5/00, A61B5/04, G01N27/27, G01N27/416	
According to International Patent Classification (IPC) or to both national classification and IPC	
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) Int. Cl. G01N33/483, A61B5/00, A61B5/04, G01N27/27, G01N27/416	
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Jitenyo Shiman Koho 1992-1996 Toroku Jitenyo Shiman Koho 1994-2001 Kokai Jitenyo Shiman Koho 1971-2001 Jitenyo Shiman Toroku Koho 1996-2001	
Electronic data base consulted during the international search (name of data base used, where practicable, search terms used)	
C. DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Relevant to claim No.
A JP 3-162663 A (Res. Dev Corp. of Japan.), 12 July, 1991 (12.07.91), (Family: none)	1-33
A JP 63-229029 A (Citizen Watch Co., Ltd.), 22 September, 1988 (22.09.88), (Family: none)	1-33
A JP 11-70090 A (Sekisui Chemical Co., Ltd.), 16 March, 1999 (16.03.99), (Family: none)	1-33
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.	
* Special categories of cited documents: "X" earlier document published after the international filing date or priority date and which is not taken into account in the search but cited to inform the applicant of the state of the art which is not considered to be of particular relevance "Y" earlier document published on or after the international filing date which may have priority rights in which is cited to establish the publication date of another document or other special reason (to be specified) "O" document referring to an oral disclosure, use, exhibition or other information not available to the public prior to the international filing date but later than the priority date claimed "A" document published prior to the international filing date but later than the priority date claimed	
Date of the actual completion of the international search 15 October, 2001 (15.10.01)	Date of mailing of the international search report 30 October, 2001 (30.10.01)
Name and mailing address of the ISA/ Japanese Patent Office	Authorized officer
Postamble No.	Telephone No.

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